



<b>ACUP ID number:</b> <i>HM-46</i>		
<b>Approval date:</b>	**	<i>GPS</i>
<b>Written by:</b>	**	<i>J. Villano</i>
<b>Revision by: (numbered accordingly)</b>	<i>08/2023</i> <i>01/2024</i>	<i>J. Villano</i>
<b>Distribution</b>	RAR	

## HM – 46: Use of Rodent Traps

**Purpose:** To prevent inhumane treatment of rodents enclosed in traps in the JHU vivaria and associated spaces like vivarium breakrooms and loading docks.

### Definitions:

Live traps: traps that will trap an animal alive and uninjured

Kill traps: traps designed to **induce the rapid death of target animals that trigger the trap**

### Personnel Responsibilities:

Animal Caretaker: To check on the rodent traps on a certain frequency identified elsewhere in this document and conduct procedures for proper euthanasia (if applicable) and disposition of the animals.

Supervisor: To ensure that staff follow proper procedures for the rodent traps.

Director and Managers, Laboratory Animal Management: Will review all deviations in this process and implement corrective action or SOP revision accordingly.

Director, Rodent Resources: Will review overall program and ensure humane vermin control

ACUC: Review and approve procedures described in this SOP.

### Materials Required:

Rodent Traps

Biohazard bags

Dead animal tags

Rubber gloves (for electric shock traps)

### Procedures:

1. Live rodent traps:
  - a. Examples of live rodent traps include Sherman traps, Ketch-alls, tin cat, and Trapper 24/7.
  - b. Check whether there is space in the trap for diet gel and ~5 feed pellets.
    - i. If so, place these provisions in the trap. **Check the traps and replace the diet gel, and if needed, the feed pellets, every 2-3 days.**
    - ii. If not, **check the traps daily.**
  - c. Euthanize any trapped rodent using the CO2 station in the animal facility. Follow relevant instructions for euthanasia.
    - i. Spray with disinfectant and wipe thoroughly the euthanasia chamber and anything that the gloved hands touched (e.g., flowmeter knob, gas tank dial) and the animal/trap came in contact with.
  - d. Re-place the live trap.
2. Kill traps:
  - a. The kill traps approved for use are electric shock traps and snap traps. Note that rodent glue traps are not allowed.
    - i. Electric shock traps
      1. Wear rubber gloves when working with this kind of traps.
      2. Such traps typically come with software management or app. Check the software/app to ensure that that the trap is recorded. Enter relevant information like location in the software/app.
      3. Catches are recorded automatically in the software/app, with an alert sent automatically to the system.
      4. Once an alert has been made, visualize the inside of the trap to ensure trapped rodent is dead.
      5. In the rare event that the rodent is not dead, do not open the trap and remove the rodent. Euthanize the rodent using the CO2 station in the animal facility. Follow relevant instructions and decontamination procedures described above.
      6. Deactivate the trap to remove the trapped rodent.
      7. **Test the trap and ensure that it functions as expected.**
      8. Re-place the trap and ensure relevant information in the software/app are accurate.
      9. **Check the traps weekly** to ensure that no rodents are trapped without the software/app not recording the event.
    - ii. Snap traps
      1. **Provision of bait**

2. **Only use when other options are not available or adequate.**
  3. **Check the traps daily.**
  4. Ensure the trapped rodent is dead.
  5. In the rare event that the rodent is not dead, euthanize using the CO2 station in the animal facility. Follow relevant instructions and decontamination procedures described above.
  6. Reset the trap, or if necessary, discard and replace.
3. Checking of traps will be done per the frequency described above for each trap type. Note that the AAALAC FAQ indicates that the monitoring frequency for live traps needs to be approved by the IACUC if it is not done daily. Entries should be made in the *Rodent Trap Monitoring Sheet* or the *Room Checklist*.
  4. Place the dead animal in a biohazard bag with a Dead Animal tag, indicating it is a wild/loose rodent and the specific location (including room number, if applicable) where it was trapped. Place the bag in a cooler and notify Rodent Resources team.
    - a. Rodent Resources Team members will collect samples for testing.
    - b. Note that the pest control vendor will submit dead trapped animals to Ross 450. Rodent Resources team members will collect samples for testing.
    - c. All efforts should be made to identify the identity of the rodent or where it came from, or at the very least, whether it is a loose laboratory rodent or a wild rodent.
  5. Report any welfare issue (e.g., dysfunctional trap) to the Director of Rodent Resources and Director/Manager of Lab Animal Management, who will then report to the pest control vendor. A decision will be made regarding necessary changes.

Reference:

1. AAALAC - FAQs. Frequency of Monitoring Rodent Traps.  
<https://www.aaalac.org/accreditation-program/faqs/#D8>

I acknowledge that I have read and understand the JHU Animal Care and Use Program document “**Use of Rodent Traps**” and I will follow this procedure. I agree to bring any deviations in this procedure to the attention of my supervisor/GPS Working Group.

\_\_\_\_\_

Name (Print) Date

\_\_\_\_\_

Signature